

Preliminary report: Zn-alpha2-glycoprotein genotype and serum levels are associated with serum lipids

Louise E. Olofsson^{a,1}, Bob Olsson^{a,*,1}, Theodore Lystig^a, Peter Jacobson^a, Margareta Jernås^a, Kajsa Sjöholm^a, Anders Gummesson^a, Lars Sjöström^a, Per Eriksson^b, Anders Hamsten^b, Laura P. Hale^c, Dag S. Thelle^d, Björn Carlsson^a, Lena M.S. Carlsson^a

^aThe Sahlgrenska Center for Cardiovascular and Metabolic Research, Department of Molecular and Clinical Medicine, Sahlgrenska Academy at University of Gothenburg, 413 45 Gothenburg, Sweden

^bAtherosclerosis Research Unit, King Gustaf V Research Institute, Karolinska Institutet, Karolinska University Hospital, 171 76 Stockholm, Sweden

^cDepartment of Pathology, Duke University Medical Center, Durham, NC 27710, USA

^dDepartment of Biostatistics, Institute of Basic Medical Science, University of Oslo, 0317 Oslo, Norway

Received 31 August 2009; accepted 2 December 2009

Abstract

Zn-alpha2-glycoprotein (ZAG) is a serum protein implicated in cancer cachexia and lipolysis. Our aim was to investigate serum levels of ZAG and polymorphisms in the ZAG gene in relation to serum lipids in man. Serum levels of ZAG correlated with serum levels of cholesterol ($P = .00088$) in healthy subjects and during weight loss ($P = .059$). The ZAG genotype was associated with total cholesterol ($P = .014$) and low-density lipoprotein cholesterol ($P = .026$) in healthy subjects, and the associations were replicated in an additional cohort ($P = .0017$ and $P = .060$, respectively). Our data indicate that ZAG plays a role in lipid metabolism.

© 2010 Elsevier Inc. All rights reserved.

1. Introduction

Zn-alpha2-glycoprotein (ZAG) is a circulating protein implicated in cancer cachexia and lipolysis [1]. Mice that are devoid of ZAG have increased body weight compared with controls [2], and ZAG has been linked to body weight in a mouse model for type 2 diabetes mellitus [3]. A recent study suggests that ZAG is a novel adipokine and that its expression in adipose tissue is down-regulated in obese subjects [1]. Thus, ZAG appears to have diverse metabolic effects. Our objective was to analyze serum levels of ZAG and polymorphisms in the ZAG gene in relation to serum lipids.

2. Methods

In the population-based Swedish Obese Subjects Reference (SOS-Ref) study, 186 healthy subjects were selected for genotyping, and in 228 subjects, serum ZAG levels were analyzed. The ZAG levels were also analyzed in 62 obese subjects before, during, and 2 weeks after a very low calorie diet (VLCD, 450 kcal/d for 16 weeks). Genotyping was also performed in 550 subjects with coronary artery disease and 550 individually age- and sex-matched controls from the Intergene study [4] and 387 patients with myocardial infarction and 387 age- and sex-matched controls from the Stockholm Coronary Atherosclerosis Risk Factor (SCARF) project [5]. The studies were approved by the local ethics committees.

Serum ZAG concentrations were determined by an in-house immunoassay [6] in the VLCD study and by enzyme-linked immunosorbent assay (BioVendor, Modrice, Czech Republic) in the SOS-Ref study. Conserved regions in the ZAG gene were identified by cross-species comparison using PipMaker (<http://bio.cse.psu.edu/pipmaker>). DNA was

* Corresponding author. Department of Molecular and Clinical Medicine, The Sahlgrenska Academy at University of Gothenburg, 413 45 Gothenburg, Sweden. Tel.: +46 31 342 32 03; fax: +46 31 41 85 27.

E-mail address: bob.olsson@medic.gu.se (B. Olsson).

¹ Both of these authors contributed equally.

sequenced with the BigDye Terminator version 3.1 cycle sequencing kit using the ABI Prism 3100 Genetic Analyzer (Applied Biosystems, Foster City, CA). The polymorphism rs4215 was genotyped with TaqMan Assays-by-Design, using the ABI Prism 7900HT Sequence Detection System.

In the SOS-Ref study, multivariate linear regression analyses were performed to test for correlation between serum levels of ZAG and lipid levels. Within-person longitudinal dependencies in the VLCD studies were addressed with generalized estimating equations to obtain adjusted standard errors. Tests of parameters using adjusted standard errors were performed using generalized Wald tests. Multivariate linear regression analyses were used to test for association between ZAG genotypes and serum lipids using an additive model where age and sex were covariates.

3. Results and discussion

In healthy subjects, serum levels of ZAG correlated with serum levels of total cholesterol ($\beta = 0.21$, $P = .00088$) and triglycerides (TG) ($\beta = 0.14$, $P = .035$) but not with low-density lipoprotein cholesterol (LDL-C) or high-density lipoprotein cholesterol (HDL-C). During diet-induced weight loss, using a combined analysis of all time points, serum levels of ZAG correlated with LDL-C ($\beta = 2.13$, $P = .033$); and there were borderline correlations to total cholesterol ($\beta = 1.88$, $P = .059$) and HDL-C ($\beta = 1.92$, $P = .055$) but no correlation with serum levels of TG. Furthermore, changes in serum levels of ZAG correlated with changes in serum total cholesterol ($\beta = 0.56$, $P < .0001$) and LDL-C ($\beta = 0.33$, $P < .0001$) in the VLCD study.

Nine polymorphisms were identified in the conserved regions of the ZAG gene, and rs4215, located in an exon, was selected for further analysis. In the SOS-Ref study, rs4215 was associated with total cholesterol and LDL-C but not with HDL-C or TG (Table 1). To replicate these findings,

rs4215 was genotyped in subjects from the Intergene and SCARF studies after exclusion of individuals that were on statin treatment or had a coronary event within 90 days before sampling [7]. In the remaining subjects ($n = 939$), polymorphism rs4215 was once more associated with serum levels of total cholesterol and LDL-C but not with HDL-C or TG (Table 1).

There are 4 very recent reports where serum levels of ZAG have been studied [8–11]. Selva et al [8] present a correlation between serum levels of ZAG and adipose tissue ZAG mRNA levels, whereas Ceperuelo-Mallafre et al [9] report the lack of correlation. This makes the relationship between serum levels of ZAG and adipose tissue-derived ZAG unclear. Yeung et al [10] report a correlation between serum levels of ZAG and TG, supporting our data from the healthy individuals. This correlation may arise from the proposed role of ZAG in adipose tissue lipolysis. Stejskal et al [11] investigated serum levels of ZAG in relation to total cholesterol levels but did not find any correlation. In our study, we found stronger relationship between serum levels of ZAG and lipids in the healthy compared with obese subjects. The reason for this is unknown; but we suspect that the difference in number of individuals analyzed may contribute, as we analyzed serum ZAG levels in 228 healthy individuals but only in 62 obese individuals. However, considering the abnormal metabolic profile in obese subjects; it is possible that there are unknown factors that affect lipid levels and disturb the normal relationship between serum ZAG and lipids. Stejskal et al [11] included both healthy subjects and subjects with metabolic syndrome, which may explain the discrepancy between the results of our studies. X-ray crystal structure of ZAG has revealed a binding site for hydrophobic ligands that has been proposed to modify the activity of ZAG [12,13]. It is possible that ligands binding to ZAG alter the properties of the protein and thereby influence ZAG interaction with other proteins or degradation of ZAG, possibly explaining the differences in relationship between serum ZAG levels and lipids in patients with different lipid profiles.

Our findings that serum levels of ZAG and cholesterol correlate in 2 separate studies and that a polymorphism in the ZAG gene is associated with circulating levels of cholesterol in 2 separate cohorts suggest that ZAG is involved in cholesterol metabolism. In obesity, more than half of the cholesterol in the body may be found in adipose tissue [14] and it has been demonstrated that the TG content in adipose tissue is paralleled by cholesterol content [15]. Furthermore, increased lipolysis leads to increased release of cholesterol from adipose tissue [16]. Therefore, it may be speculated that the correlation between serum levels of ZAG and total cholesterol is the result of the increased lipolysis of TG caused by ZAG itself and that the adipocytes adjust their cholesterol content accordingly. On the other hand, we cannot rule out that the observed relationship between ZAG and cholesterol is caused by an unrecognized direct effect of ZAG on cholesterol metabolism.

Table 1
Serum lipid levels according to polymorphism rs4215 genotype

| SOS-ref | Total cholesterol (mmol/L) | LDL-C (mmol/L) | HDL-C (mmol/L) | TG (mmol/L) |
|------------------------|-------------------------------|--------------------------|-------------------|----------------|
| CC n = 76 | 5.00 ± 0.80* | 2.96 ± 0.73 [†] | 1.66 ± 0.36 | 0.84 ± 0.27 |
| CT n = 74 | 5.29 ± 0.88 | 3.20 ± 0.82 | 1.71 ± 0.31 | 0.85 ± 0.30 |
| TT n = 36 | 5.51 ± 1.03 | 3.33 ± 0.99 | 1.79 ± 0.39 | 0.87 ± 0.25 |
| <i>Intergene/SCARF</i> | | | | |
| CC n = 352 | 5.46 ± 1.05 [‡] | 3.50 ± 0.90 [§] | 1.40 ± 0.46 | 1.45 ± 0.80 |
| CT n = 453 | 5.65 ± 1.06 | 3.59 ± 0.92 | 1.40 ± 0.46 | 1.58 ± 1.00 |
| TT n = 111 | 5.57 ± 1.02 | 3.64 ± 0.89 | 1.35 ± 0.42 | 1.50 ± 0.76 |

Analysis of CC vs not-CC using multivariate linear regression where age and sex were covariates.

* $P = .014$.

[†] $P = .026$.

[‡] $P = .0017$.

[§] $P = .060$.

In conclusion, our data indicate a link between lipids and ZAG that merits further investigation.

Acknowledgment

This work was supported by the Swedish Research Council (K2009-65X-15424-05-3, K2007-55X-11285-13-3), the Swedish Foundation for Strategic Research to Sahlgrenska Center for Cardiovascular and Metabolic Research, the Swedish federal government under the LUA/ALF agreement, the Foundations of the National Board of Health and Welfare, Torsten and Ragnar Söderberg Foundation, the Arosenius Foundation, Clas Groschinsky Foundation, Åke Wiberg Foundation, Jeansson Foundation, Tore Nilson Foundation for Medical Research, Magnus Bergvall Foundation, Wilhelm and Martina Lundgren Science Foundation, and Swedish Diabetes Foundation.

References

- [1] Bing C, Bao Y, Jenkins J, et al. Zinc-alpha2-glycoprotein, a lipid mobilizing factor, is expressed in adipocytes and is up-regulated in mice with cancer cachexia. *Proc Natl Acad Sci U S A* 2004;101:2500-5.
- [2] Rolli V, Radosavljevic M, Astier V, et al. Lipolysis is altered in MHC class I zinc-alpha(2)-glycoprotein deficient mice. *FEBS Lett* 2007;581:394-400.
- [3] Gohda T, Makita Y, Shike T, et al. Identification of epistatic interaction involved in obesity using the KK/Ta mouse as a type 2 diabetes model: is Zn-alpha2 glycoprotein-1 a candidate gene for obesity? *Diabetes* 2003;52:2175-81.
- [4] Berg CM, Lissner L, Aires N, et al. Trends in blood lipid levels, blood pressure, alcohol and smoking habits from 1985 to 2002: results from INTERGENE and GOT-MONICA. *Eur J Cardiovasc Prev Rehabil* 2005;12:115-25.
- [5] Samnegard A, Silveira A, Lundman P, et al. Serum matrix metalloproteinase-3 concentration is influenced by MMP-3 -1612 5A/6A promoter genotype and associated with myocardial infarction. *J Intern Med* 2005;258:411-9.
- [6] Hale LP, Price DT, Sanchez LM, et al. Zinc alpha-2-glycoprotein is expressed by malignant prostatic epithelium and may serve as a potential serum marker for prostate cancer. *Clin Cancer Res* 2001;7:846-53.
- [7] Brugada R, Wenger NK, Jacobson TA, et al. Changes in plasma cholesterol levels after hospitalization for acute coronary events. *Cardiology* 1996;87:194-9.
- [8] Selva DM, Lecube A, Hernandez C, et al. Lower zinc-alpha2-glycoprotein production by adipose tissue and liver in obese patients unrelated to insulin resistance. *J Clin Endocrinol Metab* 2009;94:4499-507.
- [9] Ceperuelo-Mallafre V, Naf S, Escote X, et al. Circulating and adipose tissue gene expression of zinc-{alpha}2-glycoprotein in obesity: its relationship with adipokine and lipolytic gene markers in subcutaneous and visceral fat. *J Clin Endocrinol Metab* 2009.
- [10] Yeung DC, Lam KS, Wang Y, et al. Serum zinc-alpha2-glycoprotein correlates with adiposity, triglycerides, and the key components of the metabolic syndrome in Chinese subjects. *J Clin Endocrinol Metab* 2009;94:2531-6.
- [11] Stejskal D, Karpisek M, Reutova H, et al. Determination of serum zinc-alpha-2-glycoprotein in patients with metabolic syndrome by a new ELISA. *Clin Biochem* 2008;41:313-6.
- [12] Delker SL, West Jr AP, McDermott L, et al. Crystallographic studies of ligand binding by Zn-alpha2-glycoprotein. *J Struct Biol* 2004;148:205-13.
- [13] Kennedy MW, Heikema AP, Cooper A, et al. Hydrophobic ligand binding by Zn-alpha 2-glycoprotein, a soluble fat-depleting factor related to major histocompatibility complex proteins. *J Biol Chem* 2001;276:35008-13.
- [14] Krause BR, Hartman AD. Adipose tissue and cholesterol metabolism. *J Lipid Res* 1984;25:97-110.
- [15] Kovanen PT, Nikkila EA, Miettinen TA. Regulation of cholesterol synthesis and storage in fat cells. *J Lipid Res* 1975;16:211-23.
- [16] Verghese PB, Arrese EL, Soulages JL. Stimulation of lipolysis enhances the rate of cholesterol efflux to HDL in adipocytes. *Mol Cell Biochem* 2007;302:241-8.